

Clinical profile of *FLT3*-ITD mutation in West Algerian population with acute myeloid leukemia

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ABSTRACT

Acute myeloid leukemia (AML) is a cancer of the myeloid line of blood cells, characterized by the abnormal and rapid growth of cells. The mutation of the Fms-like tyrosine kinase 3 ligand gene (*FLT3*-ITD) represents an important factor in the prognosis of AML. The objective of this study was to determine for the first time the prevalence of *FLT3*-ITD mutation in west Algerian AML patients. A total of 160 AML patients were genotyped for *FLT3*-ITD mutation by using polymerase chain reaction. *FLT3*-ITD mutation was detected in 13% of patients. Mutation rates show no significant difference in the distribution of sex and age. A positive association was found between this mutation and a higher leukocyte and blast cells counts. We also found that the M3 and M5 subtype were the commonest in the *FLT3* mutated group. This preliminary study provides first-time prevalence estimates for *FLT3*-ITD mutation in acute myeloid leukemia patients from the West region of Algeria.

Keywords: *FLT3*; ITD; Acute myeloid leukemia; Prevalence; West of Algeria

INTRODUCTION

The acute myelogenous leukemia (AML) represents approximately 80% of acute leukemias in adults and 20% of those in children. It is responsible for 1.5% of tumor-related deaths and accounts for 4 new cases /100,000 inhabitants per year [1].

Many chromosomal abnormalities involved in AML are recognized as being good prognosis markers. Commonly observed are reciprocal translocations: t (8;21), t (15;17), and pericentric inversion of chromosome 16. However, other rearrangements are associated with a poor

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prognosis such as a deletion in chromosome 5 or 7 (not specific AML type) [2, 3]. In recent years, *FLT3* has been a subject of several studies as prognostic marker in AML patients.

The Fms-like tyrosine kinase 3 (*FLT3*) gene, also named *FLK-2* (fetal liver kinase 2) or *STK-1* (human stem cell kinase 1), is located on 13q12 and consists of 24 exons [4]. It encodes a class III receptor tyrosine kinase (RTK III) and is expressed by stem cells, hematopoietic progenitors cells, brain, placenta, and liver [5]. The *FLT3* receptor plays an important role in proliferation, differentiation, and survival of hematopoietic progenitors cells, in synergy with several other growth factors [6, 7]. Hence, *FLT3* is one of the most frequently altered genes in AML. The Internal Tandem Duplications of the *FLT3* gene (*FLT3*-ITDs) were identified as the most commonly occurring mutations in patients with AML [8]. These duplications induce constitutive auto-phosphorylation of the receptor in the absence of its ligand and initiates a series of signaling events which leads to an uncontrolled cell proliferation [9, 10]. The signaling properties of a *FLT3* receptor with an ITD mutation differ from those of the wild type receptor in manner that clearly contributes to the process of leukemogenesis. STAT and FOXO transcription factors are abnormally activated in response to *FLT3*/ITD signaling. Microarray studies have identified upregulation of the *Pim1* and *Pim2* proto-oncogenes, as well as interaction with the Wnt signaling pathway in *FLT3*/ITD leukemia cells. Finally, c/EBP alpha, a transcription factor involved in myeloid differentiation, is downregulated by mutated receptors, indicative of the differentiation block that characterizes leukemia cells [10]. The frequency of *FLT3*-ITD mutation varies according to the type of leukemia: it is found preferentially in AML [8, 11, 12] but also detected at lower frequencies in myelodysplastic syndromes (MDS).

According to the literature, The *FLT3*-ITD mutation frequency varies among different ethnic populations. A relatively low frequency is identified in African populations (15-20%) [13-15] compared with other populations (20-30%) [16-18].

Indeed, mutation profiling of AML patients is part of the routine diagnostic workup for patients with de novo and recurrent AML. In Algeria, *FLT3* mutation profiling is not routinely performed in most public institutions. Therefore, there is little epidemiological data on the prevalence and heterogeneity of *FLT3* mutations and their clinical impact in Algerian AML patients. The purpose of this study is to describe the frequency of *FLT3*-ITD mutation in relation to patient demographics and specific AML classifications. Genotypic analysis of this mutation in a group of Algerian population would help and thus guide to devise precise therapeutic intervention in such patients.

MATERIALS AND METHODS

Patients: A total of 160 patients from the West of Algeria diagnosed with AML, according to WHO criteria, were included in this study. These patients were referred from hematology department of the hospital-university of Oran to cytogenetic and molecular biology laboratory of EHU d'Oran during the period of January 2014 to June 2022. Previously untreated patients were included and patients with a history of chemotherapy/radiation therapy and patients with secondary AML were excluded. Clinical data were collected by reviewing medical records. The sampling and studies were conducted after obtaining written informed consent and the study was approved by the ethics committee.

Genotyping: DNA was isolated from peripheral white blood cells by Maxwell® 16 Genomic DNA Purification Kit. The *FLT3*-ITD mutation was genotyped by polymerase chain reaction (PCR) method. The amplification was accomplished with a total 50 µl reaction mixture containing 1.5mM of MgCl₂, 200 µM of dNTP, 2.5U Taq polymerase, 0.5 µM of each primer (forward: 5'-CAATTTAGGTATGAAAGCCAGC-3'; reverse: 5'CTTTCAGCATTTTGACGG CAACC-3') and 50ng of genomic DNA. The PCR cycling conditions were as follows: 95°C for 09 min followed by 25 cycles of 95°C for 30 seconds, 57°C for 1 min, 72°C for 2 min, and a final elongation at 72°C during 10 minutes.

Statistical analysis: The demographic characteristics of AML are expressed as the mean \pm standard deviation (SD) whereas qualitative variables are presented as numbers and percentages. The comparison of qualitative data between *FLT3*-ITD⁻ and *FLT3*-ITD⁺ were evaluated using Chi-square test or Fisher's exact test .p<0.05 was considered to be of statistical significance.

RESULTS

We enrolled in this study a total of 160 AML patients from the West of Algeria. Seven samples were excluded from the analysis due to unknown genetic profiles. Out of 153 studied cases, 87 were males and 66 were women. The mean age was 37.35 ± 13.47 years. Details of demographic and clinical data are illustrated in Table 1.

Table 1: Demographics and clinical details of AML patients

Variables	Total (n=153)	Male (n=87)	Female (n=66)
Age (years)	n=138	n=78	n=60
mean \pm SD	37.35 (\pm 13.47)	38.57 \pm 13.84	35.51 \pm 13.08
Leukocytes (x10 ⁹ /L)	n=128	n=76	n=53
	3278262.4 (\pm 3191801.9)	3403371.4 (\pm 5798593.4)	3225687.4 (\pm 5560118.4)
Blast (%)	n=118	n=66	n=52
mean \pm SD	62.48 (\pm 26.84)	63.54 (\pm 27.28)	60.98 (\pm 26.23)
FAB type	153	87	66
M0	4	4	0
M1	5	2	3
M2	9	7	2
M3	8	5	3
M4	13	7	6
M5	13	10	3
M6	6	3	3
M7	1	0	1
Unknown	94	49	45

n: Number, %: Percentage, SD: Standard deviation, M0:acute myeloblastic leukemia – minimally differentiated), M1: acute myeloblastic leukemia – without maturation, M2: acute myeloblastic leukemia –with granulocytic maturation, M3: Acute promyelocytic leukemia, M4: Acute myelomonocytic leukemia, M5: Acute monocytic - monoblastic leukemia, M6: Acute erythroleukemia, M7: Acute megakaryoblastic leukemia.

The PCR amplicon of ITD mutation in the *FLT3* showed different sizes of PCR products. The 328 bp fragment indicates the size of the wild-type *FLT3* gene in the absence of ITD, while a positive result gave a band larger than 328 bp, indicating the presence of the *FLT3*-ITD mutation. A representative gel is shown in Figure 1.

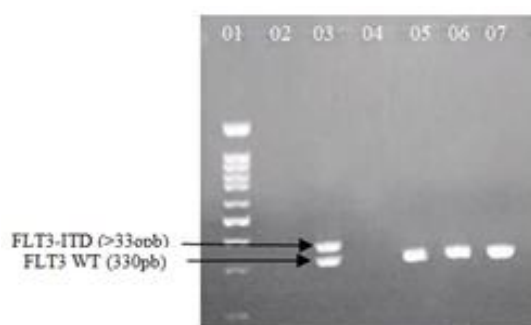


Figure 1: Representative picture of agarose gel showing wild type and *FLT3*-ITD mutant bands. Lane 01:100pb DNA Ladder, Lane02: Negative control, Lane 03: patient sample with *FLT3*-ITD mutation. Lanes 05-07: patients samples negatives for ITD mutation.

As presented in the table 2, the *FLT3*-ITD mutation was detected in 21 patients (13.7%). There was no statistically significant difference in the distribution of sex and age according to *FLT3*-ITD mutation status. However, this mutation was found to be associated with hyperleukocytosis and high blast percentage. Among FAB subtypes, *FLT3*-ITD mutation was more commonly reported within the M3, M5 subtype (Table 2).

Table 2: Distribution of *FLT3*-ITD mutations according to patient characteristics.

	Total	Wild type <i>FLT3</i>	Muted <i>FLT3</i> -ITD	P-value
	153	132 (86.2)	21 (13.7)	
Age groups n (%)				
<25	31	29 (21.9)	2 (9.5)	0.39
26-50	95	81 (61.3)	14 (66.6)	
>50	27	22 (16.7)	5 (24)	
Sex n (%)				
Male	87	75 (57)	12 (57)	0.97
Female	66	57 (43)	9 (43)	
Leukocytes mean±SD				
1-4999	n=53 2228.6(±1379.3)	n= 50 2181.4(±1376.9)	n=3 3015(±1425.5)	
5000-10000	n=15 7120.6(±1267.09)	n=14 7272.1(±1165.4)	n=1 5000	
<10000	n=60 7545767.4 (±1050860.5)	n=47 8623359.4 (±1230523.5)	n=13 61746924 (±6160469.4)	0.03
Unknown	n=25	n=21	n=4	
Blast n (%)				
1-50	60	57(43.2)	3 (14.3)	
50-100	58	43 (32.6)	15 (71.4)	0.001
Unknown	35	32 (24.2)	3 (14.3)	
FAB type n (%)				
M0	4	4 (3)	0(0)	0.41
M1	5	5 (3.8)	0(0)	0.36
M2	9	9 (6.8)	0(0)	0.21
M3	8	5 (3.8)	3 (14.3)	0.04
M4	13	10 (7.6)	3 (14.3)	0.3
M5	13	9 (6.8)	4 (19)	0.06
M6	6	5 (3.8)	1 (4.8)	0.82
M7	1	1 (0.75)	0(0)	0.69
Unknown	94	84 (63.7)	10(47.6)	/

n: Number, %: Percentage, SD: Standard deviation, M0: acute myeloblastic leukemia – minimally differentiated), M1: acute myeloblastic leukemia - without maturation, M2: acute myeloblastic leukemia – with granulocytic maturation, M3: Acute promyelocytic leukemia, M4: Acute myelomonocytic leukemia, M5: Acute monocytic - monoblastic leukemia, M6: Acute erythroleukemia, M7: Acute megakaryoblastic leukemia.

DISCUSSION

The acute myeloid leukemia is a heterogenous blood cancer which arises through the accumulation of genetic and epigenetic mutations. The *FLT3*-ITD mutation is the most frequent alteration in AML, found in approximately 25% of all cases [19].

The incidence of *FLT3*-ITD mutation in AML varies across different ethnicity. However, no data is available in Algerian patients. Currently, this is the first study to assess the prevalence of

FLT3-ITD in AML patients from Western Algeria. Furthermore, several studies have reported that the presence of this mutation is associated with a poor prognosis; thus, *FLT3*-ITD genotyping has become an essential tool to identify those who may benefit from different targeted treatment options. Therefore, this study could help to choose the optimal treatment for AML patients.

The *FLT3* is a transmembrane ligand-activated receptor tyrosine kinase. The *FLT3*-ITD mutation leads to constitutive activation of the *FLT3* Kinase which plays a key role in cell proliferation and survival of leukemic cells [7]. In fact, *FLT3*-ITD is one of the most common mutations found in AML adult patients, especially in those with a normal Karyotype. It is often associated with a high leukaemic burden which conduce a poor prognosis, and has a negative effect on the management of AML patients [16, 17]. The first generation of multitargeted tyrosine kinase inhibitors (TKIs) drug have a poor response in AML patient with *FLT3*-ITD [20, 21].

In the present work, we investigated the *FLT3*-ITD mutation status in 160 AML patients. The mean patient age at the time of testing was 37.35 ± 13.47 , ranging from 16 to 68 years. This was a youthful age compared to other populations [22-26]. These findings may be explained by the young population structure in Algeria. On the other hand, Male to female ratio was 1.3:1 which is similar to that reported by Ayachi et al. in Eastern Algeria [27] and also in most countries [28, 29]. This male predominance is poorly understood and could be attributed to the high exposure to work-related and environmental risk factors for this cancer.

In our study population, the results showed a frequency of *FLT3*-ITD mutation of 13.7%. In comparing this frequency to previous published data of different ethnic groups, this frequency is similar to those reported in Saudian [30], Egyptian [31], South African [13], Malaysian [32] and Pakistanian populations [29, 33]. On the contrary, the frequencies observed in Germany [17], Thai [18], Chinese [34], Mexican [35], Iranian [36] and Indonesian populations [37] do not agree with our findings. On the other hand, no significant differences were found between *FLT3* mutated and wild-type patients in terms of age and sex. This is in accordance with the findings of other studies [18, 31, 34]. However, Kondo et al reported a very low frequency of *FLT3*-ITD mutation in patient younger than 10 years [38].

Clinically, the results showed that AML patients with *FLT3*-ITD mutation had higher leucocytes count and an increased blast cell percentage at diagnosis compared with non mutated patients. These findings were consistent with some previous reports [39-40] and suggesting that this mutation may confer a survival advantage and increased proliferation of leukemia cells. Regarding FAB classification, this study, in accordance with others [16, 41], revealed that *FLT3*-ITD mutation occurs most commonly in M3 and M5 subtype. Otherwise, a lower frequency in M6 and M7 were reported in previous studies [16, 19, 41]. The difference in results between the current study and the previous studies can be explained by several factors such as ethnicity, patient selection criteria, genetics heterogeneity in the pathogenesis of AML and the number of patients included in different studies.

The strength of our work relies in the study of the genetic profile of third world populations in general and Algerian population in particular. In addition, this study will help to provide a platform for future diagnosis and treatment of patients with AML. However, several limitations of our study need to be mentioned: A limited number of patients and limited hematological parameters were evaluated. In addition, other biomarker such as *NPM1* mutations were not evaluated.

To the best of our knowledge, this is the first study to report the prevalence of *FLT3*-ITD mutation in our population. We identified 13% of patients harboring the mutation, which is similar to results published previously. Furthermore, the knowledge of the mutational profile of *FLT3* in a population may serve a crucial role in guiding treatment decision of AML. So, these findings need to be evaluated in a larger cohort of AML patients.

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Conflict of Interest: All authors declare that there is no conflict of interest no relevant financial or nonfinancial interests to disclose.

Authors' Contribution: Conceptualization: MSA, SD, WB; Methodology: FZM, IK, IO; Formal analysis and investigation: BW; Writing: original draft preparation: BW; Writing: review and editing: FZM; Supervision: MSA.

REFERENCES

1. Shallis RM, Wang R, Davidoff A, Ma X, Zeidan AM. Epidemiology of acute myeloid leukemia: Recent progress and enduring challenges. *Blood Rev* 2019;36:70-87.
2. Fernández de Larrea C, Kyle RA, Durie BG, Ludwig H, Usmani S, Vesole DH, Hajek R, San Miguel JF, Sezer O, Sonneveld P, Kumar SK, Mahindra A, Comenzo R, Palumbo A, Mazumber A, Anderson KC, Richardson PG, Badros AZ, Caers J, Cavo M, LeLeu X, Dimopoulos MA, Chim CS, Schots R, Noeul A, Fantl D, Mellqvist U-H, Landgren O, Chanan-Khan A, Moreau P, Fonseca R, Merlini G, Lahuerta JJ, Blade J, Orłowski RZ, Shah JJ, International Myeloma Working Group. Plasma cell leukemia: consensus statement on diagnostic requirements, response criteria and treatment recommendations by the International Myeloma Working Group. *Leukemia* 2013;27:780-791.
3. Mandal R, Bolt DM, Shah BK. Disparities in chronic myeloid leukemia survival by age, gender, and ethnicity in pre- and post-imatinib eras in the US. *Acta Oncol* 2013;52:837-841.
4. Rosnet O, Marchetto S, deLapeyriere O, Birnbaum D. Murine Flt3, a gene encoding a novel tyrosine kinase receptor of the PDGFR/CSF1R family. *Oncogene* 1991;6:1641-1650.
5. Rosnet O, Schiff C, Pébusque MJ, Marchetto S, Tonnelle C, Toiron Y, Birg F, Birnbaum D. Human FLT3/FLK2 gene: cDNA cloning and expression in hematopoietic cells. *Blood* 1993;82:1110-1119.
6. Hannum C, Culpepper J, Campbell D, McClanahan T, Zurawski S, Bazan JF, Kastelein R, Hudak S, Wagner J, Mattson J. Ligand for FLT3/FLK2 receptor tyrosine kinase regulates growth of haematopoietic stem cells and is encoded by variant RNAs. *Nature* 1994;368:643-648.
7. Gilliland DG, Griffin JD. The roles of FLT3 in hematopoiesis and leukemia. *Blood* 2002;100:1532-1542.
8. Nakao M, Yokota S, Iwai T, Kaneko H, Horiike S, Kashima K, Sonoda Y, Fujimoto T, Misawa S. Internal tandem duplication of the flt3 gene found in acute myeloid leukemia. *Leukemia* 1996;10:1911-1918.
9. Kiyoi H, Towatari M, Yokota S, Hamaguchi M, Ohno R, Saito H, Naoe T. Internal tandem duplication of the FLT3 gene is a novel modality of elongation mutation which causes constitutive activation of the product. *Leukemia* 1998;12:1333-1337.
10. Griffith J, Black J, Faerman C, Swenson L, Wynn M, Lu F, Lippke J, Saxena K. The structural basis for autoinhibition of FLT3 by the juxtamembrane domain. *Mol Cell* 2004;13:169-178.
11. Yokota S, Kiyoi H, Nakao M, Iwai T, Misawa S, Okuda T, Sonoda Y, Abe T, Kahsima K, Matsuo Y, Naoe T. Internal tandem duplication of the *FLT3* gene is preferentially seen in acute myeloid leukemia and myelodysplastic syndrome among various hematological malignancies. A study on a large series of patients and cell lines. *Leukemia* 1997;11:1605-1609.

12. Yanada M, Matsuo K, Suzuki T, Kiyoi H, Naoe T. Prognostic significance of FLT3 internal tandem duplication and tyrosine kinase domain mutations for acute myeloid leukemia: a meta-analysis. *Leukemia* 2005;19:1345-1349.
13. Marshall RC, Tlagadi A, Bronze M, Kana V, Naidoo S, Wiggill TM, Carmona SC. Lower frequency of NPM1 and FLT3-ITD mutations in a South African adult de novo AML cohort. *Int J Lab Hematol* 2014;36:656-664.
14. Kloppers JF, De Kock A, Cronjé J, van Marle AC. Molecular characterisation of NPM1 and FLT3-ITD mutations in a central South African adult de novo acute myeloid leukaemia cohort. *Afr J Lab Med* 2021;10:1363.
15. El Gammal MM, Ebid GT, Madney YM, Abo-Elazm OM, Kelany AK, Torra OS, Radich JP. Clinical effect of combined mutations in DNMT3A, FLT3-ITD, and NPM1 among Egyptian acute myeloid leukemia patients. *Clin Lymphoma Myeloma Leuk* 2019;19: e281-e290.
16. Thiede C, Steudel C, Mohr B, Schaich M, Schäkel U, Platzbecker U, Wermke M, Bornhauser M, Ritter M, Neubauer A, Ehninger G, Illmer T. Analysis of FLT3-activating mutations in 979 patients with acute myelogenous leukemia: association with FAB subtypes and identification of subgroups with poor prognosis. *Blood* 2002;99:4326-4335.
17. Fröhling S, Schlenk RF, Breitruck J, Benner A, Kreitmeier S, Tobis K, Dohner H, Dohner K, AML Study Group Ulm. Acute myeloid leukemia. Prognostic significance of activating FLT3 mutations in younger adults (16 to 60 years) with acute myeloid leukemia and normal cytogenetics: a study of the AML Study Group Ulm. *Blood* 2002;100:4372-4380.
18. Auewarakul CU, Sritana N, Limwongse C, Thongnoppakhun W, Yenchitsomanus PT. Mutations of the FLT3 gene in adult acute myeloid leukemia: determination of incidence and identification of a novel mutation in a Thai population. *Cancer Genet Cytogenet* 2005; 162:127-134.
19. Kottaridis PD, Gale RE, Frew ME, Harrison G, Langabeer SE, Belton AA, Walker H, Wheatley K, Bowen DT, Burnett AK, Goldstone AH, Linch DC. The presence of a FLT3 internal tandem duplication in patients with acute myeloid leukemia (AML) adds important prognostic information to cytogenetic risk group and response to the first cycle of chemotherapy: analysis of 854 patients from the United Kingdom Medical Research Council AML 10 and 12 trials. *Blood* 2001;98:1752-1759.
20. Stone RM, DeAngelo DJ, Klimek V, Galinsky I, Estey E, Nimer SD, Grandin W, Lebowitz D, Wang Y, Cohn P, Fox EA, Neuberg D, Clark J, Gilliland DG, Griffin JD. Patients with acute myeloid leukemia and an activating mutation in FLT3 respond to a small-molecule FLT3 tyrosine kinase inhibitor, PKC412. *Blood* 2005;105:54-60.
21. Borthakur G, Kantarjian H, Patel KP, Ravandi F, Qiao W, Faderl S, Kadia T, Luthra R, Pierce S, Cortes JE. Impact of numerical variation in FMS-like tyrosine kinase receptor 3 internal tandem duplications on clinical outcome in normal karyotype acute myelogenous leukemia. *Cancer* 2012;118:5819-5822.
22. Grimwade D. The changing paradigm of prognostic factors in acute myeloid leukaemia. *Best Pract Res Clin Haematol* 2012;25:419-425.
23. Wahlin A, Hörnsten P, Jonsson H. Remission rate and survival in acute myeloid leukemia: Impact of selection and chemotherapy. *Eur J Haematol* 1991;46:240-247.
24. Phekoo KJ, Richards MA, Møller H, Schey SA, South Thames Haematology Specialist Committee. The incidence and outcome of myeloid malignancies in 2,112 adult patients in southeast England. *Haematologica* 2006;91:1400-1404.
25. Deeg HJ. Not all patients with AML over 60 years of age should be offered early allogeneic stem cell transplantation. *Blood Adv* 2022;6:1623-1627.
26. Sultan S, Zaheer HA, Irfan SM, Ashar S. Demographic and clinical characteristics of adult acute myeloid leukemia--tertiary care experience. *Asian Pac J Cancer Prev* 2016;17:357-360.

27. Ayachi OS, Rezgoun ML, Sayitoglu M, Altindirek D, Erbilgin Y, Abadi N, Satta D. Prevalence and effect evaluation of FLT3 and NPM1 mutations in acute myeloid leukemia patients in Eastern Algeria. *Inter J Hemat Oncol* 2018;28:169-179.
28. Miller BA, Chu KC, Hankey BF, Ries LA. Cancer incidence and mortality patterns among specific Asian and Pacific Islander populations in the U.S. *cancer causes control* 2008; 19:227-256.
29. Ali A, Siddique MK, Gale RE, Shakoori AR. Frequency of FLT3/ITD Mutations in Pakistani acute myeloid leukemia patients. *Pakistan J Zool* 2013;45:495-501.
30. Elyamany G, Awad M, Fadalla K, Albalawi M, Al Shahrani M, Al Abdulaaly A. Frequency and prognostic relevance of FLT3 mutations in Saudi acute myeloid leukemia patients. *Adv Hematol* 2014;2014:141360.
31. Aly R, Shahin D, Azmy E. Prognostic significance of FLT3 internal tandem duplication in Egyptian acute myeloid leukemia and normal cytogenetics. *Comp Clin Pathol* 2012;21: 1029-1035.
32. Mat Yusoff Y, Abu Seman Z, Othman N, Kamaluddin NR, Esa E, Zulkipli NA, Abdullah J, Zakaria Z. Identification of FLT3 and NPM1 mutations in patients with acute myeloid leukaemia. *Asian Pac J Cancer Prev* 2019;20:1749-1755.
33. Ishfaq M, Malik A, Faiz M, Sheikh I, Asif M, Khan MN, Qureshi MS, Zahid S, Manan A, Arooj M, Qazi MH, Chaudhary A, Alqahtani MH, Rasool M. Molecular characterization of FLT3 mutations in acute leukemia patients in Pakistan. *Asian Pac J Cancer Prev* 2012;13: 4581-4585.
34. Xu YY, Gao L, Ding Y, Sun JZ, Wang N, Wang LL, Yu L. [Detection and clinical significance of FLT3-ITD gene mutation in patients with acute myeloid leukemia]. *Zhongguo Shi Yan Xue Ye Xue Za Zhi* 2012;20:1312-1315.
35. Cuervo-Sierra J, Jaime-Pérez JC, Martínez-Hernández RA, García-Sepúlveda RD, Sánchez-Cárdenas M, Gómez-Almaguer D, Ortíz-López R, Villarreal-Villarreal CD, Ruiz-Arguelles GJ, Ruiz-Delgado G, Lutz-Presno J, Garcés-Eisele J, Ignacio-Ibarra G, Muciño-Hernández G, Arana-Trejo RM, Jiménez-Mejia AM, Vásquez-Palacio G. Prevalence and clinical significance of *FLT3* mutation status in acute myeloid leukemia patients: A multicenter study. *Arch Med Res* 2016;47:172-179.
36. Allahyari A, Sadeghi M, Ayatollahi H, Yazdi HN, Tavakol M. Frequency of FLT3 (ITD, D835) gene mutations in acute myelogenous leukemia: a report from Northeastern Iran. *Asian Pac J Cancer Prev* 2016;17:4319-4322.
37. Notopuro PB, Jusak N, Harianto N. Detection of *flt3* gene mutations in patients with acute myeloid leukemia in surabaya, indonesia: A single-center study. *Iran J Blood Cancer* 2020; 12:54-57.
38. Kondo M, Horibe K, Takahashi Y, Matsumoto K, Fukuda M, Inaba J, Kato K, Kojima S, Matsuyama T. Prognostic value of internal tandem duplication of the *FLT3* gene in childhood acute myelogenous leukemia. *Med Pediatr Oncol* 1999;33:525-529.
39. Su L, Li W, Cui JW, Tan YH, Yang Y, Liu XL, Yu P, Hu RP, Wang LL, Gao SJ. [Correlation of NPM1, FLT3-ITD mutations with leukocyte count and myeloblasts percentage in AML patients with normal karyotype]. *Zhongguo Shi Yan Xue Ye Xue Za Zhi* 2013;21:571-575.
40. de Jonge HJ, Valk PJ, de Bont ES, Schuringa JJ, Ossenkoppele G, Vellenga E, Huls G. Prognostic impact of white blood cell count in intermediate risk acute myeloid leukemia: relevance of mutated NPM1 and FLT3-ITD. *Haematologica* 2011;96:1310-1317.
41. Schnittger S, Schoch C, Dugas M, Kern W, Staib P, Wuchter C, Löffler H, Sauerland CM, Serve H, Buchner T, Haferlach T, Hiddemann W. Analysis of FLT3 length mutations in 1003 patients with acute myeloid leukemia: correlation to cytogenetics, FAB subtype, and prognosis in the AMLCG study and usefulness as a marker for the detection of minimal residual disease. *Blood* 2002;100:59-66.